Exploring human antimicrobial antibody responses on a single B cell level

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Abstract

Analysis of monoclonal antibodies (mAbs) derived from single B cell cloning has been highly beneficial for antimicrobial immunotherapy, vaccine design, and advancing our understanding of pathogen-triggered effects on the human immunoglobulin repertoire. Sequencing of single B cells variable domains, and characterization of binding and functional activities of mAbs derived from those sequences, provides in-depth insight into not only sites of susceptibility for antibody-mediated neutralization or opsonization of the pathogen, but also on the dynamics of protective antibody evolution during infection. This information can be utilized to rapidly develop novel immunotherapies of completely human origin, and provides a roadmap for structure-based vaccine design that aims to elicit similar protective antibody responses. Here, we summarize recent aspects of the single B cell cloning approach.
Immunity from infection, natural or acquired via vaccination, requires protective memory immune responses, in many cases mediated by antibodies. The majority of microbial antigens trigger protective polyclonal antibody responses by stimulating B cells to produce protective amounts of pathogen-specific antibodies. The hosts’ ability to combat a huge variety of microorganisms is based on the immense combinatorial repertoire of circulating B cells in the periphery and organs of an individual. The calculated number of $10^{18}$ possible B cell receptor (BCR) combinations, with diversified heavy and light chain pairs, greatly exceeds the total number of B cells in an average adult (1, 2). Nonetheless, the process of affinity maturation and clonal selection results in evolution of antigen-specific antibodies within weeks or months of infection or immunization (3). Although responses to pathogens in humans and animals are polyclonal in nature, and thus heterogeneous, modern methodologies have allowed researchers to characterize the composite monoclonal antibodies (mAbs) by cloning and expressing variable domain regions from single B cells. The high-throughput nature of this approach, which often yields 50 to over 300 pathogen-specific mAbs, allows not only analysis of granular aspects of individual mAb-pathogen interactions but also provides a global view of human immune responses and how they may vary depending upon the invading pathogen. The antimicrobial human antibody repertoire can be studied by single B cell isolation and in depth repertoire sequencing; both aspects have proven to be powerful techniques for the characterization of immune responses against pathogen-specific antigens (4, 5). Following the isolation of peripheral blood mononuclear cells (PBMCs) from affected individuals, antigen-specific plasmablasts or memory B cells can be sorted by fluorescence-activated cell sorting (FACS) by labeling the cells with
a fluorescently tagged antigen, or sorting of naïve plasmablasts in acutely infected individuals. Positive hits are distributed in microtiter plates at a density of a single cell/well. The variable heavy and light chains (VH and VL, respectively) are recovered by nested RT-PCR. The recovery rate of the VH and VL is mainly based on the used primer set and the quality/quantity of the intracellular mRNA/cDNA. After isotype-specific amplification using appropriate antisense primers for the CH1 region of e.g. IgG, IgM or IgA, VH and VL regions can be grafted onto the respective constant domains. Subsequently, antibodies can be recombinantly expressed and forwarded to further characterization (4, 6, 7). Thus, recombinant mAbs can be immediately obtained without the need for B cell immortalization or fusion with myeloma partners. In general, recombinant mAbs provide an advantage in terms of reliability and reproducibility relative to polyclonal antibodies or even mAbs isolated from hybridoma cell lines. The characterization of single mAbs in combination with the respective antigen provides insights into the identification of novel mechanisms and epitopes required for successful protection against the pathogen. Those mAbs with potent protective ability against a broad range of related pathogens or strains (broadly neutralizing antibodies, bNAbes) are of great interest for antibody discovery and are promising candidates for immunotherapeutic development (4).

Recently, several previously underappreciated or insufficiently characterized pathogens triggered severe global epidemics. Such events have the capacity to result in a large number of fatal cases or chronically ill patients, or cause local public health emergencies with global implications (8). While the spread of some pathogens, e.g. *M. tuberculosis* and Dengue virus, is on-going, the epidemic spread of viruses such as LaCrosse virus, Chikungunya virus, or Zika virus was difficult to predict despite previous experiences (9). Antibody-mediated immunotherapy, consisting of single mAbs or cocktails thereof, provides an attractive platform for
initial emergency responses. Further, such epidemics have raised scientific
questions about the antigen-directed evolution of the human antibody response
during microbial exposure (10, 11). Patients who were exposed to viruses or bacteria
usually show tremendous levels of highly affinity matured immunoglobulins against
one or more antigens of the same pathogen. Studies on single B cells suggest that
pathogen-induced affinity maturation can be site specific and somatic hypermutation
is connected to the evolution of bNAbs (6, 12, 13). In this short Insight article, we
focus on recent discoveries in the field of antimicrobial antibody responses that were
achieved by single B cell analysis.

Insights into B cell expansion and seroconversion from single B cell sorting

Pathogen-mediated B cell receptor crosslinking triggers the activation of
antigen-specific B cells (ASBCs) that subsequently undergo massive cell
proliferation. During the course of an infection, the overall number of ASBCs in the
periphery mainly depends on the elapsed time from exposure (6, 14, 15). Several
studies recorded incremental changes in plasmablast counts to identify the ASBC-
specific subpopulation by a time course-dependent flow cytometric analysis (14, 15).
After acute dengue virus (DENV) infection, the number of virus-specific plasmablasts
peaks within the first week after antigen exposure (15, 16). For an acute DENV
infection, numbers of DENV-specific CD19^+ CD3^− CD20^−/low CD38^{hi} CD27^{hi} plasmablasts rise on average from 0% to 47%
(15). In total numbers, DENV infections boost the average basal level of 3.7 × 10^5
plasmablasts to more than 1 × 10^6 plasmablasts per milliliter of blood (15). This result
affirms the high immunogenicity of DENV in comparison to (for example) yellow fever
virus, another flavivirus, that shows the highest peak at day 7 and day 11 with an
overall of <7% of reactive plasmablasts, respectively (15). These temporarily high numbers of B cells might induce high rates of acute and covalent seroconversion in individuals over a long time (6). For instance, a small population of patients that were vaccinated with the *M. tuberculosis* vaccine BCG displayed unusually high serum levels against the *M. tuberculosis* derived antigen lipoarabinomannan (AM) even >10 years after vaccination (12). Flow cytometry analysis of AM-reactive B cells reveals a very low number of antigen-specific B cells that might be responsible for the production of significant amounts of monoclonal antibodies that are efficient enough to provide protection.

For a large number of approved vaccines (e.g. Measles, Mumps, Rubella vaccine, yellow fever), antibody titers stay consistently protective in a large number of patients over the course of several years, while other vaccines require booster immunization (e.g. Tetanus) (17-19). Antigen-mediated boosting triggers the differentiation of previously primed B cells and the generation of antigen-specific plasmablasts (20). A fraction of these plasmablasts then migrate into specialized bone marrow niches and contribute as long-lived plasma cells to the maintenance of the serum immunoglobulin levels (20, 21). Technological advances in mass spectrometry and single B cell cloning have facilitated a deeper understanding into the serological memory of vaccinations. To correlate circulating or resting long-lived plasma cells with their actual ability for immunoglobulin expression levels in the serum, NGS of the V gene repertoire can be paired with high-resolution liquid chromatography tandem MS (MS/MS) (19, 22). To elicit a correlation between existing plasmablasts and actual immunoglobulin expression, the peripheral repertoire of anti-tetanus antibodies was investigated before and after tetanus (TT) booster vaccination (19). Deep sequencing of the TT+ serum IgG repertoire of two donors recorded between 80-100 different TT+ specific clonotypes after the booster
vaccination. Hereby, three clonotypes represented >40% of the overall TT⁺-specific immunoglobulin levels in the serum. The remaining clonotypes were present at frequencies less than 0.5% each. Comparison of the clonotypic repertoire vs. the expressed serum immunoglobulin repertoire revealed that only a small fraction of plasmablast clonotypes or memory B cells encode for antibodies that contribute nine months after booster vaccination to the Ig serum. This suggests that a large population of booster responsive TT⁺ plasmablasts were not part of the long-lived serological memory (19). The high quality of selection for specific B cells is an important process that mainly facilitates an efficient immune response.

**Affinity maturation during antigen exposure**

The goal of many vaccinations is to mimic protective aspects of pathogen-induced B cell response. Over the time course of multiple vaccinations, antibodies are selected and affinity matured to improve specificity. However, hypervariable viruses such as influenza or HIV-1 that mutate in high frequency have been particularly recalcitrant to vaccine development. Efficient protection against HIV-1 and influenza virus likely requires the induction of bNAbs which, even in actively viremic individuals, may be exceedingly rare or not present (23). Isolation of bNAbs by single sorting has revealed that a large portion of broadly neutralizing influenza, HIV-1 or HCV antibodies is derived from the VH1-69 germline segment (23). The unexpectedly high representation of bNAbs from the VH1-69 germline segment implies that viral infections can alter adaptive immune responses and bias or deregulate typical antibody germline usage. Interestingly, VH1-69-derived antibodies show a high percentage of somatic hypermutations based on a critical polymorphism (23). It can be presumed that those specific B cells derive from germinal centers with
a high mutagenic activity (24). The reconstruction of genealogical trees of multimember clones from influenza virus exposed and vaccinated individuals revealed mutation sites within CDRs of isolated anti-hemagglutinin bnAbs that seem to be specifically favored during the process of bnAb evolution and are important sites for affinity maturation (23). Pappas et al. discovered five antibodies that were clonally related, based on the conserved HCDR residues R30, A52, and Y98. Back mutation of these residues to the germline sequence did not abolish binding to the virus. However, it was shown that the germline sequence of the antibody lead to the reduction of neutralization of other H1 strains. This implies that site-specific evolution of certain residues during antibody development is essential for the development of broadly neutralizing activity. Site-specific affinity maturation was also shown to be important in broadening the specificity for other influenza antibodies (13). The antibody 3I14 derived from IGHV3-30 is able to bind the H3 and H1 subtypes but not H5 in its germline configuration (D94). Interestingly, germline reversion of the somatic hypermutation D94N significantly restores the binding to H5 underlining that certain residues are important for broad antiviral protection (13).

Understanding the site-specific SHMs that give rise to bNAbs in HIV-1 and influenza virus infections might be therefore provide an opportunity for immunogen engineering in a way to properly guide affinity maturation toward protective bnAb responses by vaccination. For the HIV-1 bnAb PGT121 and related members, it was previously shown that the maturation of these narrowly related bnAbs may be mainly based on the specific interaction or avoidance of steric clashes with a high mannose glycan on the HIV-1 trimer (25). It was demonstrated that SHMs on some of the PGT121-like mAbs promote CDR configurations to avoid non-productive interactions with glycosylated residue N137, while in other cases, SHMs themselves contribute to direct interactions with the same glycan. Removal of glycosylated N137 from gp120
increased the binding affinity of all PGT121-related mAbs by significant amounts suggesting that the presence of the glycan essentially shapes the maturation process of the antibody (25). In general, highly mutated HIV-1 antibodies have a significant higher potential to protect due to a re-occuring exposure of the molecules to the antigen over time. It was shown that not only the virus, but also the bNAbs evolve over time in long-term HIV-1 infection (26). Time-dependent sequencing of antibodies from the CH103 lineage paired with neutralization and binding studies demonstrated that unmutated antibodies can evolve to broadly neutralizing molecules within 34 months (26). Due to chronic exposure of the virus, SHM rates incrementally change from 0% to 17% and show that the mostly evolved antibody CH103 is neutralizing more than 50% of the tested HIV-1 pseudoviruses in comparison to clonally related antibodies with a lower rate of mutation (26). Interestingly, the maturation process focused on sites that were only minimally affected by viral escape mutations. This result implies a continual “host-virus” battle that favors antibodies that suppress a broad array of viruses.

For the elicitation of bNAbs by immunization, it is likely important that the vaccine is able to prime germline B-cell receptors, later inducing a mature bNAb lineage. This was achieved by Jardine et al. who remodeled the CD4 binding site on gp120 to target VRC01-class bNAb germline precursors, then evaluated this immunogen as a multivalent immunonanoparticle-based vaccine eOD-GT6 (27). After crosslinking of a specific class of BCRs by the resurfaced CD4 binding site, antigen-specific germline B cells proliferated and matured towards broadly neutralizing VRC01-class B cells. VRC01-class bnAbs neutralized HIV-1 as they mimic CD4 by binding to gp120 (27). To identify the precursor VRC01-B cells, the affinity of the immunonanoparticles towards VRC01-class derived antibodies was optimized by yeast display panning against germline-derived and mature VRC01
antibodies (28). The evolved immunogen eOD-GT8 was then used as a probe for epitope-specific B cell sorting. Highly stringent B cell sorting from 15 donors revealed that only 1 in 2.4 million naïve B cells were derived from the VRC01 lineage (28). This provides an explanation about the rarity of these antibodies but also demonstrates the precision of single B cell sorting with an optimized antigen.

Altogether, smart vaccine design can mainly improve the induction of broadly specific antibodies. However, sophisticated analysis of the evolution of single B cell populations and a detailed structural analysis have to be performed to design a rational concept for broadly protective vaccines (29).

Isolation of antigen-specific molecules for immunotherapy from single B cells

In some cases, the application of vaccines is inappropriate or not manageable due to circumstances such as severe acute infections or cross-reactivity with other pathogens. In these cases, immunotherapies consisting of one or more mAbs provide an attractive therapeutic alternative. Furthermore, even for pathogens where an effective vaccine exists, it may be desirable to have accompanying immunotherapies for use in cases where the vaccine is inaccessible or not adopted. Rapid development of immunotherapies for newly emerging pathogens, or analysis of antibody repertoires, are depending on highly sophisticated technology platforms such as single B cell sorting or hybridoma technology. In acute cases, it is important to utilize the most rapid direct method for the isolation of potent human antibodies to generate therapeutic molecules for clinical applications - even within a few weeks (4). In comparison to hybridoma technology or immunization of transgenic animals (e.g. with humanized Ig loci), there are significant advantages of B cell sorting. Hybridoma technology and murine immunization requires long-term screening and animal
handling. Furthermore, it is important to consider that murine antibody responses might differ from human patterns. Murine antibodies were also shown to trigger immunogenic responses, provide less stability and hamper downstream Fc effector functions. Based on all of these pitfalls, our lab and others use antigen-dependent B-cell sorting for the isolation of a variety of different pathogen specific molecules for a straightforward mAb isolation within a few weeks (4, 30). For the detection of antigen-specific B cell populations, the antigen must be either labeled by fluorescently labeled streptavidin or by another fluorescent tag. For our work we observed that the selection and the careful design of the antigen is critical to the quality of the isolated antibodies. Random biotinylation methods can either decrease the number of specific molecules or block important functional binding sites that lead to the exclusion of potential top hits. The configuration of the sorting antigen (e.g. trimeric, monomeric) is equally important. A large number of potent molecules for e.g. Dengue or Zika Virus rely on quaternary epitopes that can be only fished out by mimicking the natural architecture of the viral surface (31, 32).

A successful isolation of a large number of human antibodies was performed for an Ebola virus (EBOV) patient with elevated glycoprotein (GP_{EBOV}) serum responses. It was shown that 1-3% of memory B cells were GP_{EBOV} reactive three month after infection (4). By using high-throughput single B cell sorting, Bornholdt et al. isolated 349 antibodies against different neutralization relevant epitopes of GP_{EBOV} from the survivor of the 2014 Ebola outbreak (4). Thereby, analysis of GP_{EBOV} reactive VH and VL revealed a widely distributed usage of clonal lineages meaning that the isolated antibodies massively differ in V/J combinations, CDR3 loop lengths and CDR3 sequence homologies (4). This result contrasts with studies isolating influenza or HIV-1 specific antibodies that seem to favor particular germline lineages depending upon epitope (14, 33). Downstream characterization of antigen-specific
antibodies such as epitope mapping and binding affinity studies unravel novel epitopes and provide information about the immunogenicity or mechanisms of action of the antigen binding sites. Most of the human antigen responses against \( \text{GP}_{\text{EBOV}} \) are directed against well-characterized GP epitopes such as the glycan cap or the GP1/GP2 interface (4). However, one of most potent \( \text{GP}_{\text{EBOV}} \) antibodies from this study, ADI-15758, binds proximally to the viral membrane – a so far less characterized epitope that was presumably hardly accessible – and demonstrated efficient protection against lethal doses of EBOV in mice. Based on the visualization of three ADI-15758 Fab-molecules complexed with three \( \text{GP}_{\text{EBOV}} \), it was assumed that the HR2 region of the pre-GP might already exist as a three-helix bundle. This shows that information from single B cell isolation and epitope mapping provide valuable predictions towards the structural function of the viral glycoprotein.

Similar studies were also performed for the isolation of broadly neutralizing Dengue virus antibodies. Dengue virus is the leading arthropod-transmitted viral disease in the world with approximately 400 million human infections per year (34). The mature, prefusion glycoprotein E exists as a head-to-tail dimer organized into rafts with icosahedral geometry on the viral particle. Each E subunit contains three domains, DI, DII, and DIII. The post-fusion E structure is a trimer with DIII and the stem region significantly relocated relative to DI and DII, so as to bring the host and viral membranes into proximity to facilitate viral membrane fusion. One promising class of protective molecules are described as antibodies that bind the E dimer epitope (EDE) on the E protein efficiently neutralizing multiple DENV serotypes (34). Dejnirattisai et al. sorted plasmablasts from seven acutely infected patients and isolated 145 antibodies by single B cell cloning (35). Antibodies to EDE were shown to be highly crossreactive against multiple DENV strains (34). Epitope mapping by alanine-scanning reveals that EDEs is either an epitope that is contained within an E
dimer or is shared by adjacent E dimers on the viral surface (3,4). For the isolated EDE2 antibody 747(4)B7 it was shown that EDE dependent neutralization does not necessarily depend on the crosslinking of two adjacent dimers. Cryo-electron microscopy of the Fab 747(4)B7 showed that EDE was only bound at an intradimer epitope but not across dimers. Other EDE antibodies such as EDE1 C8 however are using the binding across dimers to efficiently cross-neutralize Dengue and Zika viruses in a highly efficient way (35).

Despite the very efficient generation of antibodies, there are many challenges for single B cell sorting. Because of the unbiased nature of this procedure, not all isolated antibodies express very well due to a certain V gene usage. Another pitfall is linked to the appropriate isolation of the correctly assembled constant region of the antibodies. Some applications require the right combination between the isotype and the variable region to trigger the appropriate downstream effector functions. So far, recent polymerase technologies limit the isolation of antibodies only to the variable regions based on the polymerases’ limited half life during excessive PCR cycle numbers. However, since protein engineering is rapidly evolving, this problem might be solved in the near future. A final limitation is that the B-cell sorting method does not correlate with secreted mAbs, and thus the individual composite mAbs may not reflect the serum response. Nonetheless, all mentioned studies show that high throughput screening of PBMCs can lead to an efficient generation of new molecules for immunotherapy and to a better understanding of the pathogen-triggered immune response. The isolation of monoclonal human antibodies is a powerful tool to map out important binding epitopes and unravel neutralization mechanisms for a large number of different pathogens in a short time.
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Figure 1: Workflow for the isolation of monoclonal antibodies by antigen-dependent single B lymphocyte sorting (Abbreviations: PBMC: Peripheral blood mononuclear cells; RHP: random hexamer primers; PCR: Polymerase chain reaction; VH: variable domain of the heavy chain; VL: variable domain of the light chain; HC: Heavy chain; LC: Light chain; mAb: monoclonal antibody; BLI: Biolayer interferometry; ELISA: Enzyme-linked-immunosorbent assay).