Serodiagnosis of Recently Acquired Toxoplasma gondii Infection Using an Enzyme-Linked Immunosorbent Assay with a Combination of Recombinant Antigens

SHULI LI,1,2 GINA GALVAN,1 FAUSTO G. ARAUJO,1 YASUHIRO SUZUKI,1,2 JACK S. REMINGTON,1,2,6 AND STEPHEN PARMLEY†1

Department of Immunology and Infectious Diseases, Research Institute, Palo Alto Medical Foundation, Palo Alto, California 94301,1 and Division of Infectious Diseases and Geographic Medicine, Department of Medicine, Stanford University School of Medicine, Stanford, California 943052

Received 10 March 2000/Returned for modification 8 May 2000/Accepted 14 June 2000

An enzyme-linked immunosorbent assay (ELISA) using four recombinant antigens of Toxoplasma gondii (rP22, rP25, rP29, and rP35) was used in an attempt to differentiate pregnant women with toxoplasma serologic profiles (TSPs) indicative of recently acquired infections (acute profile) from those with TSPs indicative of infections acquired in the distant past (chronic profile). In general, immunoglobulin G antibodies in sera from women with the acute profile reacted more strongly with the recombinant antigens than did those in sera from women with the chronic profile. However, reactivities differed significantly between antigens that reacted with a single serum and between sera that reacted with a single antigen. Because of these variations, we employed a combination of the four antigens in an ELISA (Comb-ELISA) and evaluated its ability to distinguish pregnant women with the acute profile from those with the chronic profile. Eighteen of 20 (90%) sera from acute-profile women were positive in the Comb-ELISA, whereas 69 of 70 (98.6%) sera from the chronic-profile women were negative. Thus, the Comb-ELISA may be useful for diagnosis of toxoplasmosis in pregnant women and for differentiation between recently acquired infections and infections acquired in the more distant past.

MATERIALS AND METHODS

Serum samples. Sera were provided by the Toxoplasma Serology Laboratory of the Research Institute, Palo Alto Medical Foundation, Palo Alto, Calif. Because the main objective of the study was to determine whether a recently acquired infection with T. gondii could be differentiated from an infection acquired in the distant past using a single serum sample from a pregnant woman, all sera used in the study were from pregnant women. The sera were divided into three groups: group I sera were from 26 women with TSPs consistent with recently acquired T. gondii infections (acute profile), group II sera were from 70 women with TSPs consistent with infections acquired in the distant past (chronic profile), and group III sera were from 20 women who were seronegative for T. gondii antibodies. The 20 women in group III were healthy pregnant women with no reported illness, and their ages were in the same range as those of the other groups. The serologic profile of each sample was based on the results of the following serologic tests performed in the Toxoplasma Serology Laboratory: Sabin-Feldman dye test (DT), IgM ELISA, IgA ELISA, and AC/HS test (14, 15, 28). The results of these tests comprise the TSP (14). Sera from women in group I had high DT titers, positive IgM and IgA ELISA results, and acute patterns in the AC/HS test. Sera from women in group II had low DT titers, negative IgM and IgA ELISA results, and chronic patterns in the AC/HS test. The classification of acute or chronic profile was based on the results of the TSP in combination with the individual’s clinical history (14).

Ten sera from each group were used to determine the reactivity of each serum with individual antigens and the optimal concentration of each antigen. Ten sera from each of the three groups were randomly chosen and coded, and then they were used in a blind study to determine the effectiveness of the ELISA with combined recombinant antigens (Comb-ELISA) to differentiate sera from the three groups. Thirteen of these 30 sera were used further in an experiment to determine the reproducibility of the Comb-ELISA.

Infections, including human immunodeficiency virus infection, other than with
Production and purification of recombinant proteins. For expression of recombinant pMAL-P22, -P25, or -P29 (rP22, rP25, rP29) or nonrecombinant MBP, Escherichia coli strain JM101 (Stratagene, La Jolla, Calif.), transformed with pMAL-c2 (New England Biolabs), was grown in LB medium supplemented with 50 μg of ampicillin per ml and 0.2% glucose at 37°C overnight. Two-liter culture flasks containing 400 ml of LB medium supplemented with 50 μg of ampicillin per ml and 0.2% glucose were inoculated with 10 ml of the overnight culture. The cultures were grown at 37°C with vigorous shaking until the optical density at 600 nm reached 1.0. Isopropyl-β-D-thiogalactoside (IPTG) was added to a final concentration of 0.5 mM, and growth was continued for 4 h at 37°C. The cells were centrifuged at 4,000 × g for 20 min at 4°C. The pellets were resuspended in 50 ml of column buffer (20 mM Tris-HCl [pH 7.4], 1 mM EDTA, 200 μg of MBP per ml) at 4°C. Sodium glutamate was added until the pH was 2.5 for subsequent resuspension. The cells were disrupted by sonication in an ice-water bath in pulses of 15 s each. The sonicated sample was centrifuged at 9,000 × g for 40 min at 4°C. The fusion proteins were purified from the supernatants (crude extracts) by affinity chromatography according to the manufacturer's instructions (New England Biolabs). Briefly, the crude extracts were diluted 1:5 with column buffer and applied to a 10-ml amylose resin column at a flow rate of 1 ml/min. After extensive washes with column buffer, fusion proteins were eluted with column buffer containing 10 mM maltose.

For expression of recombinant pGEX-S-1-P35 (rP35) or GST protein, E. coli strain BL21 (Pharmacia Biotech, Piscataway, N.J.), transformed with the recombinant plasmids, was grown in LB medium supplemented with 50 μg of ampicillin per ml at 37°C overnight. Two-liter culture flasks containing 400 ml of LB medium supplemented with 50 μg of ampicillin per ml were inoculated with 5 ml samples of the overnight cultures. The cultures were grown at 30°C with vigorous shaking until the optical density at 600 nm reached 0.8 to 1.0. IPTG was added to a final concentration of 1.0 mM, and growth was continued for 4 h at 38°C. The cells were pelleted at 7,700 × g and 4°C for 10 min. The pellets were resuspended in 20 ml of 1× Tris-buffered saline (TBS)-1% Triton X-100. Samples were frozen at −20°C overnight. Resuspended cell pellets were sonicated as described above and centrifuged at 12,000 × g for 10 min at 4°C. Recombinant proteins were purified from the supernatants (crude extracts) using a batch purification protocol (Pharmacia Biotech). Briefly, 50% glutathione-Sepharose 4B slurry was prepared according to the protocol. Two milliliters of slurry was added to crude extracts, and the slurry was incubated with gentle agitation at room temperature for 60 min. The pellets were washed twice with 50 ml of 1× TBS-1% Triton X-100 followed by two washes with 50 ml of 1× TBS. Recombinant proteins were eluted with glutathione elution buffer (10 mM glutathione, 50 mM Tris-HCl [pH 8.0]).

ELISA with individual recombinant antigens for demonstration of IgG antibodies. The optimal concentration of the recombinant antigens for ELISA was determined using the checkerboard method (4). Three sera from each group were used. Different concentrations of each recombinant antigen in 0.1 ml of carbonate buffer were used to coat the wells of microtiter plates. The optimal concentrations, which provided the greatest difference between the absorbencies noted with sera from group I and from group II, were 3 μg/ml for rP22, rP25, rP29, and 5 μg/ml for rP35. These concentrations, in a volume of 100 μl of 0.05 M carbonate buffer, pH 9.4, were used to examine the antibodies individually. Control wells were coated with 3 μg of MBP per ml or with 5 μg of GST per ml in 100 μl of carbonate buffer. Coating was performed at 4°C overnight. Thereafter, plates were washed with PBS-Tween and postcoated with 200 μl of 3% BSA per well in PBS at 37°C for 2 h. After washing, 100 μl of serum diluted 1:50 in 3% BSA in PBS was applied to each well. Plates were incubated at 37°C for 1 h and then washed, and 100 μl of horseradish peroxidase-conjugated goat anti-human IgG (Caltag Laboratories) diluted 1:8,000 was added to each well. After 1 h of incubation at 37°C, the plates were washed, 100 μl of 0.03% o-phenylenediamine in H2O2 was added to each well, and then the plates were incubated at room temperature for 10 min. The test was read using an automatic ELISA reader (Dynatech Laboratories, Chantilly, Va.). Each sample was run in duplicate. Results were determined for each serum by calculating the mean value of the absorbency readings for duplicate wells. The final results were reported after the MBP or GST control protein reading was subtracted from the sample reading for each of the recombinant antigens.

Comb-ELISA for detection of IgG antibodies. To evaluate the ability of the Comb-ELISA to distinguish recently infected women from women who acquired the infection in the distant past, the wells of the microtiter plates were coated with 100 μl of carbonate buffer containing 3 μg each of rP22, rP25, and rP29 per ml and 5 μg of rP35 per ml. Control wells were coated with 100 μl of carbonate buffer containing 9 μg of MBP per ml plus 5 μg of GST per ml. Thereafter, the plates were incubated with sera and conjugate as described above.

RESULTS

Reactivity of T. gondii IgG antibodies in immunoblots with recombinant antigens. Three pools of sera, each consisting of five sera from group I, II, or III, were examined in immunoblots to determine the reactivities of IgG antibodies to T. gondii recombinant antigens, and nonrecombinant control proteins. Whereas IgG antibodies in group I and group II sera reacted strongly with T. gondii, IgG antibodies in group III sera did not react with T. gondii or with any of the recombinant antigens (data not shown). Figure 1 shows that IgG antibodies in the five pooled group I sera reacted strongly with each of the recombinant antigens, whereas IgG antibodies in group II sera re-
acted weakly with rP22 and rP29 and did not react with rP25 or rP35. IgG antibodies in each serum pool did not react with the MBP or GST control. Gel electrophoresis to determine the molecular weights of the recombinant antigens revealed that the positions of the bands corresponding to each recombinant antigen on the gel were consistent with the molecular weights calculated from the sizes of the gene fragments encoding each antigen (data not shown).

**Reactivities of IgG antibodies to T. gondii in ELISA with individual recombinant antigens.** Ten sera from each group were examined. None of the sera from group III had a positive reading with any of the recombinant antigens after the absorbency readings of the control proteins were subtracted (data not shown). All 10 sera from group I (Fig. 2) and 6 of 10 sera from group II (Fig. 3) had absorbency readings indicative of a positive result with each recombinant antigen. The degree of antibody reactivity within the same serum differed for each antigen. To differentiate group I from group II sera, a cutoff value for each recombinant antigen was established as the mean plus 2 standard deviations of the absorbency readings obtained with the 10 sera from group II. Using these cutoff values (Fig. 4), two sera from group I (numbers 4 and 9) were negative with rP22, two (numbers 5 and 9) were negative with rP25 and rP29, and four (numbers 3, 5, 7, and 9) were negative with rP35. One serum from group II had readings slightly above the cutoff value with rP29 (number 10) and rP35 (number 5).

**Reactivities of IgG antibodies to T. gondii in the Comb-ELISA.** The 20 sera from groups I and II used in the ELISA with individual recombinant antigens were used in the experiments to evaluate the Comb-ELISA (Fig. 5). The cutoff value was calculated as described above, using results obtained with sera from group II. Whereas none of the sera from this group had readings above the cutoff value, 9 of 10 sera from group I had readings above the cutoff value. It was interesting that sera 3, 4, 5, and 7 from group I, which were negative with one or
more antigens when tested with individual recombinant antigens (Fig. 4), were positive when tested in the Comb-ELISA. Only serum 9 remained negative.

To better differentiate group I from group II sera, we examined 60 samples from the latter group in the Comb-ELISA. The results for these sera, combined with the results for the 10 sera from the same group as described above, were used to establish a cutoff value that was more representative of the reactivities of sera from group II (chronic profile) in the Comb-ELISA. The cutoff value was 0.065 and represented the reactivities of 70 sera from group II in the Comb-ELISA. Only 2 (2.8%) of the 70 group II sera had absorbency readings above the cutoff value (data not shown). In contrast, 18 of 20 (90%) sera from group I (acute profile) had readings above the established cutoff value (Fig. 6).

In a previous study employing ELISA with a single recombinant antigen (rP35) (13), it was found that IgG antibodies to rP35 were not detected in six sera with TSPs consistent with the acute profile. When the same sera were examined in the Comb-ELISA, two of the six sera gave readings suggestive of the acute profile (Fig. 7).

In the blind study (Fig. 8), 100% of sera from women who were not infected had absorbency readings far below the cutoff value (Fig. 8, group III sera). Of 10 sera with TSPs compatible with past infection, one had an absorbency reading minimally above the cutoff value, and the remaining 9 had absorbency readings well below the cutoff value (Fig. 8, group II sera). Of 10 sera with TSPs compatible with recent infection, 2 had absorbency readings below the cutoff value, and the remaining 8 had readings well above the cutoff value (Fig. 8, group I sera).

Statistical analysis (by analysis of variance) of the results of the reproducibility experiment revealed no significant variation among either the means ($P = 0.77$) or the standard deviations ($P = 0.48$).

**DISCUSSION**

Because *T. gondii* can be transmitted from a recently infected mother to her fetus, a rapid and accurate diagnosis of the infection is critical for establishing proper clinical care. Frequently, only a single sample of serum is available for determination of whether the infection was acquired recently or in the distant past. In the present study, we compared the
reactivities of IgG antibodies to four recombinant antigens of *T. gondii* in sera from pregnant women with acute and chronic TSPs. IgG antibodies in sera from women with acute TSPs reacted to the four recombinant antigens more strongly than did antibodies in sera from those with chronic TSPs. Immunoblots also revealed remarkable differences in the reactivities of IgG antibodies from these two groups of women with the individual recombinant antigens.

Significant variations in the reactivities of IgG antibodies were noted when sera from the pregnant women were tested in an ELISA with individual recombinant antigens. When the reactivities of IgG antibodies from each serum were analyzed, the differences were more evident. For example, one serum reacted with rP25, rP29, and rP35, but not with rP22; another reacted only with rP22. These variations indicate the difficulty in developing a serologic test using only a single recombinant antigen.

Because of the problems encountered when a single recombinant antigen was used to distinguish women with acute TSPs from those with chronic TSPs during gestation, we attempted to differentiate these two groups of individuals using the Comb-ELISA. The Comb-ELISA proved effective in differentiating acute from chronic TSPs; the sensitivity and specificity were 90 and 97%, respectively.

It was interesting that five sera with acute TSPs were negative when tested with the individual recombinant antigens. However, four of these sera were positive when tested using the Comb-ELISA. Moreover, in a previous study in which rP35 ELISA was employed (13), six sera from pregnant women with TSPs indicative of recent infection were negative. When these six sera were tested in the Comb-ELISA, two were positive for recently acquired infection.

By combining several recombinant antigens that present multiple different epitopes, the probability of detecting *T. gondii* antibodies during different stages of the infection will likely be increased (2, 10, 12).

A number of reports (17, 22, 25) have described the successful use of recombinant antigens for detection of antibodies to *T. gondii*. In some of them (12, 25), the antigens were used to attempt to distinguish between acute and chronic infections. However, it is difficult to compare the results of these studies because the criteria for acute and chronic infections vary.
among investigators. For most, the presence of specific IgG and IgM antibodies was sufficient to define serum samples as being from acutely infected individuals, while the absence of specific IgM antibodies was sufficient to define serum samples as being from chronically infected individuals. However, because of the demonstrated persistence of IgM antibodies during the chronic stage of infection with T. gondii (14, 26), this criterion alone cannot be used to distinguish acute from chronic infections (23). In the United States, only a single serum sample is usually available with which to determine if the infection was acquired during pregnancy. This problem was the impetus for our studies on such sera to attempt to differentiate recent from past infections with T. gondii. Our results suggest that combinations of recombinant antigens will be useful for serologic diagnosis of toxoplasmosis in pregnant women and for differentiation between a recently acquired infection and one acquired in the distant past.

ACKNOWLEDGMENTS

We thank Greg Maine and Sean Nowland for their critical discussions and Xiulan Zhou for technical assistance.

This work was supported by Public Health Service grant AI04717.

REFERENCES


Hidalgo, A. Fleuret, and P. Ambroise-Thomas. 1998. Determination of anti-
Toxoplasma gondii immunoglobulin G avidity: adaptation to the Vidas sys-
sis of Toxoplasma gondii antigens recognized by human sera obtained be-
1987. Toxoplasma gondii antigens recognized by sequential samples of serum
1931.
Remington. 1990. Cloning, expression, and cDNA sequence of surface anti-
using a recombinant form of the dense granule antigen GRA6 in an enzyme-
antibodies for diagnosis of acute congenital and acquired toxoplasmosis.
26. Wilson, M., J. S. Remington, C. Clavet, G. Varney, C. Press, D. Ware, and
commercial kits for detection of human immunoglobulin M antibodies to
27. Wong, S. Y., M.-P. Hajdu, R. Ramirez, P. Thulliez, R. McLeod, and J. S.
Remington. 1993. Role of specific immunoglobulin E in diagnosis of acute
Infect. Dis. 18:853–862.