Can an Antibiotic (Macrolide) Prevent \textit{Chlamydia pneumoniae}-Induced Atherosclerosis in a Rabbit Model? \\

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There is increasing data implicating \textit{Chlamydia pneumoniae} in the pathogenesis of atherosclerosis, and antibiotics may theoretically be useful to prevent secondary vascular complications. Three groups of New Zealand White specific-pathogen-free rabbits, fed cholesterol-free chow, were inoculated via the nasopharynx on three occasions, 2 weeks apart, with \textit{C. pneumoniae}. Group I \((n = 23)\) rabbits were untreated; group II \((n = 24)\) rabbits were treated with azithromycin at 30 mg/kg of body weight daily for 3 days and then once every 6 days, starting 5 days after first inoculation and continuing until sacrifice (early treatment); and group III \((n = 24)\) rabbits were treated with the same dose of azithromycin but initiated 2 weeks after the last inoculation. All animals were sacrificed at 10 to 11 weeks after initial inoculation and examined for signs of atherosclerosis of the aorta. Eight (34.8\%) untreated rabbits developed early signs of atherosclerosis, whereas only one (4.2\%) in the early-treatment group had such signs \((P = 0.02)\). However, eight rabbits (33.3\%) of the delayed-treatment group had atherosclerotic changes of the aorta and no significant reduction compared to untreated rabbits. Early treatment of \textit{C. pneumoniae}-infected rabbits with azithromycin was highly effective (87\%) in preventing atherosclerotic changes, but delayed treatment was ineffective. It is possible that longer or more aggressive antibiotic treatment may be needed to reverse preformed lesions or that antibiotics may not be of value once lesions have formed.

\textit{Chlamydia pneumoniae}, a respiratory pathogen in humans, has been associated with coronary heart and carotid artery diseases in many seroepidemiological studies (6, 17, 20, 21, 24, 28, 29, 32, 33). Moreover, the organism can be detected in fatty streaks and atheromatous plaques from the aorta and coronary, carotid, and femoral-popliteal arteries but not in normal vessels (1, 2, 4, 13–15, 31). Viable organisms have also been recovered from atheromas of carotid and coronary arteries (12, 18, 26).

Further evidence of the role of \textit{C. pneumoniae} in atherogenesis is supported by animal models with rabbits reported by us and others (7, 16, 23). Recently, two small pilot clinical trials have suggested that newer macrolides (which are effective in vitro against \textit{C. pneumoniae}) may decrease secondary cardiovascular events after myocardial infarction or unstable angina (9, 10).

It is important, however, to determine whether antimicrobial agents are of value in preventing or reversing early aortic lesions that are inducible in the rabbit model with \textit{C. pneumoniae}. These studies may provide insight into and guidance on the potential dosages and duration of therapy for future clinical trials in humans.

**MATERIALS AND METHODS** \\

This study was approved by the Animal Care Committee of our institution, and guidelines for care of the animals were strictly adhered to.

**Animals.** One-month-old, male, New Zealand White, specific-pathogen-free rabbits, fed standard chow (no cholesterol supplementation), were used in the experiments. Three groups of animals \((24\) per group) were inoculated via the nasopharynx with a small catheter on three separate occasions. 2 weeks apart, with \textit{C. pneumoniae}. Group I rabbits were untreated (controls); group II received azithromycin at 30 mg/kg of body weight daily for 3 days and then once every 6 days, starting after the 5th day post-initial inoculation and continuing until sacrifice; and group III received the same dosage schedule of azithromycin but start 2 weeks after the last inoculation. Animals were sacrificed and aortas were examined for signs of atherosclerosis 10 to 11 weeks after initial inoculation.

\textit{C. pneumoniae} strains and inoculum. TWAR strain AR-39 (Seattle, Wash.) and ATCC strain VR 1310 (originally respiratory isolates) were used in the studies. The organisms were grown in HEp-2 cells (27). Infected cells were harvested with sterile glass beads and ultrasonic disruption after 72 h. Cell culture-grown organisms were partially purified by one cycle each of low- and high-speed centrifugation, resuspended in sucrose-phosphate-glutamic acid buffer, and frozen in 1.0-ml aliquots at \(-70^\circ\text{C}\). Inoculum preparations were adjusted to contain \(1.5 \times 10^7\) to \(2.6 \times 10^7\) inclusion-forming units of \textit{C. pneumoniae}. Contamination by \textit{Chlamydia trachomatis}, \textit{Chlamydia psittaci}, or \textit{Mycoplasma} species was excluded by analysis with PCR genus- and species-specific primers (26) and monoclonal antibody staining.

**Serology.** Antibodies (immunoglobulin G [IgG]) to \textit{C. pneumoniae} were measured by the microimmunofluorescence test (MRL Diagnostics, Cypress, Calif.). The IgG serum antibody fractions were measured by using fluorescein-isothiocyanate-conjugated goat anti-rabbit IgG (Jackson Laboratories, West Grove, Pa.). Blood was obtained from the earlobe marginal veins on the day of sacrifice. Our previous studies of 36 rabbits showed no detectable \textit{C. pneumoniae} antibodies at baseline before inoculation. IgM and IgA antibodies were not tested because anti-rabbit IgM and IgA conjugates were not available commercially. The \textit{C. pneumoniae} antibodies were tested in all three groups by screening at a 1:16 dilution against purified elementary bodies of \textit{C. pneumoniae} (AR-39). All serum samples positive at 1:16 had their titers determined in twofold dilutions to endpoint.

**Pathology.** At sacrifice, the entire aorta from the ascending aorta to the iliac bifurcation was removed and cleansed of any fat. The aorta was split longitudinally, and sections were taken from the arch and descending and abdominal aorta.

All specimens obtained were fixed in 10\% buffered formalin, processed, and paraffin embedded. Staining with hematoxin and eosin stain and an elastic stain (Movat’s pentachrome) was performed on sections for histological examination. Aortic lesions were graded histologically, modified from the work of Daley et al. (5), as follows: grade I, early fatty streaks; grade II, advanced fatty streaks; grade III, spindle cell lesion, defined as consisting of spindle-shaped (smooth muscle) cells and

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other products; grade IV, advanced atheromatous lesion, defined as the presence of a core containing pools of extracellular lipid and/or necrotic debris and fibrous cap. Calcification of a fibromuscular lesion without a lipid core was also classified as grade IV. None of the infected rabbits developed the classic grade IV lesion with a lipid core, but rather they had fibromuscular lesions with calcification.

Myxoid lesions were defined as isolated thickening of the intima-media with ground substance or myxoid-like material which stained green with Movat’s stain but had no other features of atherosclerosis as defined. These lesions were usually associated with fragmentation and separation of the elastic fibers in the media. Periaortitis represented focal areas of accumulation of inflammatory mononuclear cells between the outer wall of the aorta and the adventitia. These cells consisted of macrophages and T and B lymphocytes, shown by immunohistochemical staining with specific monoclonal antibodies.

Immunohistochemical study. Immunohistochemical staining (14) for C. pneumoniae antigen was performed on paraffin-embedded sections by the labelled strept avidin-biotin-peroxidase method (3) with the Histo-Stain kit of Zymed. The antibodies used included C. pneumoniae-specific monoclonal antibody RR-402 and Chlamydia genus-specific antibody CF-2 (Washington Research Foundation, Seattle), and a second C. pneumoniae-specific monoclonal antibody, Chlamydia Cel Pn (Cell Lab). In selected cases, assays with factor VIII and smooth muscle actin (DAKO, Carpinteria, Calif.) and T cells, B cells, and macrophage markers specific for rabbits (Serotec) were performed on adjacent sections to identify cells of endothelium, smooth muscle, T and B cells, and macrophages, respectively.

A negative control consisting of phosphate-buffered saline instead of primary antibodies was used in each run. Positive controls for C. pneumoniae included positive lung and spleen cells from our previous study (7), and cell pellets of the C. pneumoniae inoculum prepared in HEp-2 cells were fixed in formalin and embedded in paraffin.

Data analysis. The prevalence of atherosclerotic lesions in the treated groups was compared to that in the untreated group by chi-square analysis or Fisher’s exact test.

RESULTS

One of the control untreated animals was sacrificed early because of an accident and was not included in the analysis. Eight (34.8%) of the 23 untreated rabbits had early signs of patchy atherosclerosis (predominantly grade III lesions). Another five (21.7%) rabbits had myxoid-like material accumulating between the intima-media but no other changes suggestive of atherosclerosis. Eight (34.8%) rabbits demonstrated focal mononuclear cell infiltration in the adventitia of the abdominal aorta classified as periaortitis (Fig. 1).

The effects of the azithromycin treatment on the various pathological entities are summarized in Table 1. Early treatment with azithromycin was highly effective in preventing early atherosclerotic lesions (87%) and periaortitis (100%) but had no effect on the myxoid-like changes. Delayed treatment was ineffective in reducing or reversing atherosclerotic lesions and myxoid-like changes but was highly effective in reducing periaortitis (100%). The results of the immunostaining for C. pneumoniae antigen are summarized in Table 2. Azithromycin did...
not decrease the prevalence of detectable antigen between the untreated controls and early-treated rabbits, but *C. pneumoniae* was detected more frequently in the delayed-treatment group than in the early-treatment group.

**Serology.** All animals in the control untreated group had antibodies to *C. pneumoniae* at a >1:16 dilution (range, 1:32 to 1,024) and one (4.2%) of the animals in the early-treated group had positive serology of 1:16, but antibodies in the rest were undetectable (<1:16). Six (25%) of the animals in the delayed-treatment group had undetectable antibodies, but the remaining 18 (75%) had antibodies ranging from a 1:16 to a 1:256 dilution. None of the rabbits with undetectable antibodies had evidence of aortic lesions.

**DISCUSSION**

We have previously shown that *C. pneumoniae* infection in the rabbit can induce early lesions of atherosclerosis de novo, whereas these changes are not seen with sham-infected controls or *Mycoplasma pneumoniae*-infected rabbits (20, 29). This study clearly demonstrates that early treatment with azithromycin, 5 days after initial infection, was highly effective (87%) in preventing atherosclerosis-like lesions or periaortitis. The significance of the periaortitis and myxoid-like changes is not clear, and we cannot explain the reason why the myxoid lesions were not decreased with treatment but periaortitis was prevented. Myxoid or ground substance (glycosaminoglycan) accumulation is commonly seen in human atherosclerotic lesions (preatheroma and more advanced lesions) but may also be present in Marfan’s syndrome and vascular inflammation (30, 34). This is believed to be a response of smooth muscle cells to inflammatory stimuli, with increased production of glycosaminoglycan. Since we did not analyze the biochemical nature of our lesions, we may not be able to reverse preformed atherosclerotic lesions but still effective (100%) in preventing periaortitis. Treatment with the macrolide did not significantly decrease the prevalence of *C. pneumoniae* antigen in the aorta. Detectable antigen in the aorta was more frequently localized to atherosclerosis-like lesions or areas of periaortitis but could be seen in areas without histological changes. Although not all lesions with atherosclerotic changes demonstrated *C. pneumoniae* antigen, this could be related to sampling at a specific microscopy level. However, we could not determine whether there were viable organisms, as *C. pneumoniae* is extremely difficult to recover by culture from rabbits and can be recovered only from the respiratory tract within 2 to 3 days of inoculation (22).

The results of the *C. pneumoniae* serology assay showed that early treatment was largely effective in abrogating the antibody response (and was correlated with lack of aortic lesions), compared to delayed treatment. This would suggest that the antigens detectable in the early-treated group represent nonviable bacteria or existence in a latent stage without stimulation of the immune system.

In a study of different design, Muhlestein et al. (23) were able to show that azithromycin at 30 mg/kg intramuscularly daily for 1 week and then twice weekly for another 6 weeks (starting 3 days after final triple inoculation of *C. pneumoniae*) was able to reverse the enhanced intimal thickening of the aortas in the rabbits fed a 0.25% cholesterol-enriched chow. Our study did not use a cholesterol-enriched diet and therefore, it looked at the de novo effect of *C. pneumoniae* infection on induction of early atherosclerosis. Moreover, in our study the animals were treated by oral gavage with less frequent dosing.

Azithromycin, a newer macrolide antibiotic of the azalide subclass, has in vitro activity against *C. pneumoniae* similar to that of erythromycin with a MIC against 90% of strains of 0.125 to 0.25 μg/ml (11). Although the absolute bioavailability is only 37%, this drug penetrates intracellularly in a wide variety of tissues (including macrophages where *C. pneumoniae* may reside), with an intracellular concentration of 200 to 500 times the levels in plasma or extracellular levels (25). The dosage used in our study was designed to mimic the human life of 63 h in humans, allowing once-a-week dosing, and a slightly shorter half-life in rabbits (24a). It is possible that with more frequent dosing and longer therapy we may have greater efficacy with the delayed treatment. But it is also possible that we may not be able to reverse preformed atherosclerotic lesions, as we previously showed that grade I to III lesions can be formed within a month of *C. pneumoniae* infection in rabbits (7). Since in the delayed-treatment group the rabbits received

### TABLE 1. Effects of early and delayed treatment with azithromycin on aortic lesions

<table>
<thead>
<tr>
<th>Condition</th>
<th>No treatment* (n = 23)</th>
<th>Early treatment* (n = 24)</th>
<th>Delayed treatment* (n = 24)</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>Atherosclerosis</td>
<td>8 (34.8)</td>
<td>1 (4.2)</td>
<td>8 (33.3)</td>
<td>0.02</td>
</tr>
<tr>
<td>Myxoid-like change</td>
<td>5 (21.7)</td>
<td>8 (33.3)</td>
<td>4 (16.7)</td>
<td>NS</td>
</tr>
<tr>
<td>Periaortitis</td>
<td>5 (21.7)</td>
<td>0</td>
<td>21 (87.5)</td>
<td>&lt;0.02</td>
</tr>
</tbody>
</table>

* Number of rabbits (percent) showing condition.

**TABLE 2. *C. pneumoniae* antigen detection after treatment with azithromycin**

<table>
<thead>
<tr>
<th>Treatment group (n)</th>
<th>No. of aortic segments with <em>C. pneumoniae</em> antigen/total no. (%)</th>
<th>No. of segments (%) showing degree:</th>
<th>No. of animals (%) with <em>C. pneumoniae</em> antigen</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>1+ 10</td>
<td>2+ 10</td>
</tr>
<tr>
<td>Control (23)</td>
<td>11/69 (15.9)</td>
<td>10</td>
<td>8 (34.8)</td>
</tr>
<tr>
<td>Early azithromycin (24)</td>
<td>14/72 (19.4)</td>
<td>12</td>
<td>9 (37.5)</td>
</tr>
<tr>
<td>Delayed azithromycin (24)</td>
<td>45/72 (62.5)</td>
<td>39</td>
<td>21 (87.5)</td>
</tr>
</tbody>
</table>

* 1+, 5 to 10 cells positive in one focus. 2+, two or more foci positive.
at least 4 to 5 weeks of therapy without benefit, and the normal lifespan of a rabbit is 7 to 8 years, 10-fold less than that of humans, this would suggest that short-term therapy in humans may not be effective and that clinical trials should use a longer duration, of more than 40 to 50 weeks of treatment, to assess the clinical benefit of azithromycin in vascular complications of atherosclerosis.

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REFERENCES


25. Pfizer Inc. In-house data. Personal communication.


