Patients with multiple myeloma and other B cell disorders respond poorly to pneumococcal vaccination. Vaccine responsiveness is commonly determined by measuring pneumococcal serotype-specific antibodies by enzyme-linked immunosorbent assay (ELISA), by a functional opsonophagocytosis assay (OPA), or by both assays. We compared the two methods in vaccinated elderly patients with multiple myeloma, Waldenstrom’s macroglobulinemia, and monoclonal gammopathy of undetermined significance (MGUS). Postvaccination sera from 45 patients (n = 15 from each patient group) and 15 control subjects were analyzed by multiplexed OPA for pneumococcal serotypes 4, 6B, 14, and 23F, and the results were compared to IgG and IgM antibody titers measured by ELISA. While there were significant correlations between pneumococcal OPA and IgG titers for all serotypes among the control subjects (correlation coefficients [r] between 0.51 and 0.85), no significant correlations were seen for any of the investigated serotypes in the myeloma group (r = -0.08 to 0.21) or in the group with Waldenstrom’s macroglobulinemia (borderline significant correlations for 2 of 4 serotypes). The MGUS group resembled the control group by having good agreement between the two test methods for 3 of 4 serotypes (r = 0.53 to 0.80). Pneumococcal postvaccination IgM titers were very low in the myeloma patients compared to the other groups and did not correlate with the OPA results. To summarize, our data indicate that ELISA measurements may overestimate antipneumococcal immunity in elderly subjects with B cell malignancies and that a functional antibody test should be used specifically for myeloma and Waldenstrom’s macroglobulinemia patients.
cently been refined, and new serotype-specific correlates for both ELISA and functional antibodies in infants have been proposed (17). The pneumococcal serotype-specific IgG ELISA has been the standard method due to its relative simplicity and cost-effectiveness, but it has its drawbacks, not least the capture of potentially nonfunctional, cross-reacting antibodies, although this problem has largely been overcome by adsorption of cell wall polysaccharide and heterologous pneumococcal polysaccharide (22F) (18). The assessment of the functionality of pneumococcal antibodies by a biological opsonophagocytosis assay (OPA) is more complicated and time-consuming but measures the functional antibody titers. It encompasses mixing various dilutions of patient sera with phagocytic HL60 cells, complement, and live pneumococci and has the actual number of surviving bacteria as the readout. It has been multiplexed for several serotypes and is regarded as the gold standard for the evaluation of pneumococcal vaccine responses (19).

Determination of antipneumococcal IgG levels by means of later generations of serotype-specific ELISA has been shown to correlate well with opsonic activity in children but more poorly in adults, in particular, in the elderly population (19, 20). The knowledge regarding the performance of the ELISA for determination of pneumococcal vaccine efficacy in high-risk groups such as immunocompromised patients is limited. Patients with B cell malignancies have not been investigated in this aspect. In our previous study, we found unexpectedly high antipneumococcal antibody levels among some myeloma patients, possibly indicative of the presence of cross-reactive, nonfunctional antibodies (14). We therefore wished to assess the reliability of the serotype-specific pneumococcal ELISA in patients with multiple myeloma and related B cell disorders, who, due to their susceptibility to pneumococcal infections, constitute a target group for vaccination. To this end, we compared the results from serotype-specific IgG ELISA with those of a standardized OPA. Since the antibodies that mediate killing-type OPA may be of any isotype, we also investigated the postvaccination serum levels of IgM and their correlation to OPA titers.

MATERIALS AND METHODS

Study population. In an earlier study, we investigated pneumococcal vaccine responses in 56 patients aged ≥60 years with MM (n = 24), WM (n = 15), or MGUS (n = 17) and 20 control subjects without hematological disorders but within the same age range, recruited at the Department of Hematology, Uddevalla Hospital, Uddevalla, Sweden, between May 2008 and March 2009 (14). The study persons were randomized to receive a single dose (0.5 ml) of either the 7-valent conjugated pneumococcal vaccine (PCV7) (Prevenar; Pfizer) or the 23-valent polysaccharide pneumococcal vaccine (PPV) (Pneumo 23; Sanofi Pasteur). Serum samples were collected before and 4 to 8 weeks after vaccination and were stored at −20°C until analyzed. The study was approved by the Regional Ethics Committee in Göteborg, and written informed consent was obtained from all patients.

From the above-mentioned study population, 15 patients from each disease group and 15 control subjects were randomly selected for reanalysis of their postvaccination sera by 4-fold multiplexed OPA and pneumococcal serotype-specific IgM ELISA. Both assays were performed at the UCL ICH laboratory by adapting a previously described IgG ELISA protocol (http://www.vaccine.uab.edu/ELISA%20Protocol.pdf) (23). Each serum was tested in duplicate.

Statistical analyses. Geometric means with 95% confidence intervals were calculated for ELISA and OPA titer. Since no significant titers differences were found, these samples were not excluded from the study.

IgM pneumococcal ELISA. Serum was assayed for IgM antibodies to pneumococcal serotypes 4, 6B, 14, and 23F at the UCL ICH laboratory adapting a previously described IgG ELISA protocol (http://www.vaccine.uab.edu/ELISA%20Protocol.pdf) (23).

RESULTS

Study population. Patient characteristics are presented in Table 1. More than half (55%) of the study persons were female. The median ages were significantly higher in the MM and WM groups (80 and 75 years, respectively) than in the MGUS and the control study groups (71 and 68 years, respectively; P < 0.05). Eleven of the MM patients (73%) and only one of the WM patients were on active cancer treatment during the study (Table 1).

Serology. Since no significant differences in OPA IgG or IgM antibody titers/concentrations for any of the four serotypes were found between the subgroups of patients vaccinated with PCV7 or PPV, the data were pooled for the correlation analyses (data not shown).

OPA titers. All three patient groups had lower antipneumococcal OPA titers than the control subjects (Fig. 1). This was significant for all four pneumococcal serotypes in the group of WM patients, for three (serotypes 4, 6B, and 14) in the MM group, and for two (serotypes 6B and 14) among the MGUS patients (Fig. 1). The distribution of the unreportable OPA results among the study
groups was as follows: 2 results from the MM group, 5 results from the WM group, 3 results from the MGUS group, and 2 results from control subjects. Eight of the 12 unreportable results were single individual samples and individual serotypes, and 4 results were from 2 WM patients, each with two serotypes affected (serotypes 6B and 23F, respectively). There was no obvious skew regarding the unreported OPA results in favor of certain serotypes or patient groups.

**Correlations between ELISA IgG and OPA titers.** The correlations between antipneumococcal IgG concentrations and functional antibody titers as measured by OPA are presented in Fig. 2. While there were significant correlations between ELISA concentrations and OPA titers for all four pneumococcal serotypes among the control subjects (correlation coefficients \([r]\) between 0.51 and 0.85), no correlations were found for any of the investigated serotypes in the myeloma group (\([r]\) between 0.18 and 0.21). Correlations were significant for three serotypes (serotypes 4, 14, and 23F) among the MGUS patients (\([r]\) between 0.53 and 0.80) and almost reached significance for serotypes 4 and 23F in the WM group (\([r]\) 0.54 and 0.53; \([P]\) 0.06) (Fig. 2).

**ELISA IgM concentrations for pneumococci.** IgM antibody titers were low among the MM patients for all four pneumococcal serotypes, both in relation to the vaccine-induced IgG titers and in comparison with the IgM responses seen in the other three study groups (\([P]\) 0.001 for MM patients versus control subjects for serotypes 4, 6B, and 14; \([P]\) 0.05 for serotype 23F) (Fig. 3). The control subjects, patients with WM, and MGUS patients had comparable IgM titers after vaccination (Fig. 3).

**Correlations between ELISA IgM and OPA titers.** The correlations between ELISA IgM and OPA titers were generally poor and inconsistent. Significant correlations were found only for WM patients regarding serotype 6B and for the control subjects regarding serotypes 4 and 14 (Table 2). No significant correlation coefficients were seen for either MM or MGUS patients (Table 2).

**DISCUSSION**

In a previous study of pneumococcal vaccine responses in elderly patients with multiple myeloma, Waldenstrom’s macroglobulinemia, and monoclonal gammopathy of undetermined significance, we suspected falsely high ELISA IgG titers, in particular, among

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**TABLE 1 Study group characteristics**

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<thead>
<tr>
<th>Subject characteristic</th>
<th>Value for study group</th>
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<tbody>
<tr>
<td></td>
<td>Multiple myeloma</td>
</tr>
<tr>
<td>(n)</td>
<td>15</td>
</tr>
<tr>
<td>Median age, yrs (range)</td>
<td>80 (65–88)</td>
</tr>
<tr>
<td>Female sex, (n)</td>
<td>7</td>
</tr>
<tr>
<td>M-protein median, g/liter (range)</td>
<td>24 (0.7–44)</td>
</tr>
</tbody>
</table>

Receiving indicated treatment regimen(s), \(n\)
- Melphalan + prednisone: 4
- Cyclophosphamide + dexamethasone: 4
- Pulse steroids: 1
- Thalidomide: 1
- Bortezomib: 1
- Fludarabin: 1

Receiving ongoing antibiotic treatment, \(n\)
- 3

Given pneumococcal vaccine, \(n\)
- PCV7\(^b\): 8
- PPV23\(^c\): 7

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\(^a\) Prevaccination levels of monoclonal (M-)protein in serum.
\(^b\) PCV7, 7-valent pneumococcal conjugate vaccine.
\(^c\) PPV23, 23-valent pneumococcal polysaccharide vaccine.
We therefore wished to assess the ELISA results by comparison with OPA findings. Excellent correlation between IgG concentrations determined by ELISA and OPA titers were seen for the control subjects for all four serotypes. In contrast, multiple myeloma patients showed no positive correlation between IgG and OPA titers for the four serotypes studied. High ELISA IgG titers with no functional activity were noted in some MM patients. Of the remaining two groups of patients studied, the MGUS patients resembled the control subjects by having good correlations between their IgG and OPA titers for 3 of 4 of the serotypes while the Waldenstrom’s macroglobulinemia patients had intermediate correlations.

Discrepancies in the outcome of pneumococcal ELISA and OPA such as those described in this study have previously been noted for other immunocompromised patients with increased risk of pneumococcal infections, for example, for HIV patients and renal transplant patients. In contrast, our results differ from those of a study of allogeneic stem cell-transplanted patients with hematological malignancies, where significant correlations between antipneumococcal IgG and OPA titers were found for all seven investigated pneumococcal serotypes. Of note is that the patients in that study (mean age, 37 years) were younger than ours and had different hematological diagnoses and thus may have been able to mount better IgG responses. Also, the correlations were modest, with \( r \) values ranging from 0.48 to 0.76.

Age is an important factor in evaluations of pneumococcal vaccine responses. Previous studies have shown poorer functional antibody responses after vaccination with PPV among elderly adults than among younger adults. The 13-valent conjugate pneumococcal vaccine (PCV13), which was launched after the initiation of this study, has been shown to prevent invasive pneumococcal disease and community-acquired pneumonia in

![FIG 2 Correlations between antipneumococcal IgG titers (ELISA) and opsonophagocytic antibody titers (OPA) after pneumococcal vaccination. (A) Serotype 4. (B) Serotype 6B. (C) Serotype 14. (D) Serotype 23F. MM, multiple myeloma; WM, Waldenstrom’s macroglobulinemia; MGUS, monoclonal gammopathy of unknown significance. Diamonds indicate persons vaccinated with 7-valent conjugated pneumococcal vaccine (PCV7); circles indicate persons vaccinated with 23-valent pneumococcal polysaccharide vaccine (PPV). Note that the values on the x and y axes differ between the different pneumococcal serotypes (A to D). \( r \) and \( P \) values were calculated by the Spearman rank sum test.](http://cvi.asm.org/)

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adults >65 years of age (28); however, vaccine efficacy declined with increasing age (29). Since the control subjects and the MGUS patients in this study were significantly younger than the MM and WM patients, this might have influenced our results, with poorer OPA concentrations in the more diseased (and older) study groups (the MM and WM groups).

The weak correlations between ELISA-determined IgG concentrations and OPA titers among MM and WM patients in the present study could also have been affected by the advanced age of these study persons, which would be in accordance with previous studies (19, 20). Romero-Steiner et al. found poorer correlations between postvaccination IgG antibody concentrations and OPA titers in a group of elderly adults (mean age, 85 years) than in younger adults (mean age, 37 years). This was explained by lower antibody avidity with respect to capsular polysaccharides among the elderly (20) and was corroborated by showing that transfer of human serum with a high OPA titer and high avidity protected mice in an experimental model of pneumococcal infection, whereas serum with low avidity and a low OPA titer did not do so despite containing the same concentration of antipneumococcal IgG (20).

In general, higher antipneumococcal IgG levels are required to achieve the same killing capacity in elderly adults as in younger subjects (20, 26, 27, 30). Besides correlating with the differences in avidity (20), high age has been linked to loss of oligoclonality in response to pneumococcal antigens, reduced function of phagocytic cells, and decreased levels of antipneumococcal IgM as a consequence of the presence of deficient IgM memory B cells (26, 31, 32). In the present study, IgM concentrations were markedly reduced in the myeloma patients compared to the other study groups. In contrast, serum IgM levels were relatively high among the WM patients; however, a significant correlation with OPA titers was seen for only 1 of 4 tested serotypes. Our results showing poor or no correlation between IgM and OPA titers for all study groups indicate that lack of IgM was not the main reason for poor OPA function among our study subjects.

A possible reason for the poor correlations between the ELISA and OPA analyses among our disease groups might be interference of the M-proteins with the ELISA analysis. This has been shown for various laboratory tests such as analyses of blood counts and electrolytes but also for immunological tests such as analysis of antistreptococcal antibodies (33). Unfortunately, we had too little serum left to measure the M-protein levels in the postvaccination samples and could not correctly test this hypothesis. However, this should not be the sole explanation since the correlations between IgG and OPA titers were nearly normal among the MGUS patients, who had prevaccination levels of M-protein that were almost as high as in the WM group (a median of 11 g/liter and a range of 0.5 to 28 g/liter compared to a median of 15 g/liter and a range of 3 to 28 g/liter among the WM patients). The OPA responses might also be affected by deficient cellular immunity, which has been shown to be more pronounced in MM and WM patients than in MGUS patients (8).

The major limitation of this study was the small series of patients. The age differences between 2 of the 3 patient groups (MM and WM) and the control subjects may also have affected the results. Nevertheless, an evaluation of methods for measuring

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**TABLE 2** Correlation of ELISA IgM level and opsonophagocytosis assay results for four pneumococcal serotypes and four study groups

<table>
<thead>
<tr>
<th>Study group</th>
<th>r value, by pneumococcal serotype</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Serotype 4</td>
</tr>
<tr>
<td>MM</td>
<td>0.39</td>
</tr>
<tr>
<td>WM</td>
<td>0.54</td>
</tr>
<tr>
<td>MGUS</td>
<td>−0.16</td>
</tr>
<tr>
<td>Controls</td>
<td>0.56*</td>
</tr>
</tbody>
</table>

* Each group contained 15 postvaccination serum samples. MM, multiple myeloma; WM, Waldenström’s macroglobulinemia; MGUS, monoclonal gammopathy of unknown significance. *, P < 0.05; ***, P < 0.001 (Spearman rank sum test).
pneumococcal vaccine responses has not to our knowledge previously been done for patients with B cell malignancies and related hematological disorders. Since patients with these diagnoses have a high incidence of pneumococcal infections and thus are target groups of pneumococcal vaccination, it should be of interest to find a sufficiently reliable method of measuring their vaccine responses.

Our main finding was that pneumococcal antibody measurement by ELISA in vaccinated elderly patients with multiple myeloma and Waldenström’s macroglobulinemia did not correlate with antibody titers determined by the multiplex OPA. We suggest that ELISA measurements may overestimate antipneumococcal immunity in patients with B cell malignancies and propose the use of a functional antibody assay, in particular, for multiple myeloma and Waldenström’s macroglobulinemia patients.

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