A major impediment to tuberculosis (TB) vaccine development is the lack of reliable correlates of immune protection or biomarkers that would predict vaccine efficacy. Gamma interferon (IFN-γ) produced by CD4+ T cells and, recently, multifunctional CD4+ T cells secreting IFN-γ, tumor necrosis factor (TNF), and interleukin-2 (IL-2) have been used in vaccine studies as a measurable immune parameter, reflecting activity of a vaccine and potentially predicting protection. However, accumulating experimental evidence suggests that host resistance against Mycobacterium tuberculosis infection is independent of IFN-γ and TNF secretion from CD4+ T cells. Furthermore, the booster vaccine MVA85A, despite generating a high level of multifunctional CD4+ T cell response in the host, failed to confer enhanced protection in vaccinated subjects. These findings suggest the need for identifying reliable correlates of protection to determine the efficacy of TB vaccine candidates. This article focuses on alternative pathways that mediate *M. tuberculosis* control and their potential for serving as markers of protection. The review also discusses the significance of investigating the natural human immune response to *M. tuberculosis* to identify the correlates of protection in vaccination.

**The Problem: Disconnect between Polyfunctional T Cells and Vaccine Efficacy**

Partial or complete gamma interferon (IFN-γ) receptor deficiency in humans leads to disseminated nontuberculous mycobacterial (NTM) infections or BCGosis, and mice deficient in IFN-γ exhibit growth and virulence (4). These modified strains of *M. tuberculosis* are under preclinical assessments. MVA85A, the first booster vaccine candidate to complete an efficacy trial since BCG, did not provide significantly higher protection (5), despite exhibiting a significantly higher level of antigen-specific T cell responses during preclinical development (6). This setback in TB vaccine development has reinforced the importance of revisiting and revising our understanding of host immune components that can serve as reliable markers of protection in vaccine-mediated immunity. In this article, we first discuss the growing literature which indicates that there is a disconnect between polyfunctional T cells and vaccine efficacy. Next, we deliberate on whether immune cells other than CD4+ T cells potentially correlate with protection and the emerging concept that the innate compartment has memory-like facets. We also discuss the relevance of clinical studies focused on tracking the natural course of human immune response to *M. tuberculosis* and large-scale data analysis tools to identify correlates of protection. Our aim for this review is to draw attention to mechanisms beyond conventional memory T cells and cytokines. There are exhaustive reviews on host immunity, memory T cells, and cytokines in TB, and therefore, these topics have not been reviewed.
impared control of bacterial growth and dissemination (7–10). Furthermore, IFN-γ production is depressed in whole-blood cultures from advanced TB patients (11). Together, these findings led to the assumption that the robust production of IFN-γ is a strong correlate of protection and thus a useful readout for testing immunogenicity of TB vaccine candidates. Subsequently, work from several studies revealed that IFN-γ is not a reliable measure of protection against M. tuberculosis (12, 13). Although systemic production of IFN-γ by M. tuberculosis-stimulated peripheral blood mononuclear cells (PBMC) from patients with moderate and far-advanced pulmonary TB is depressed, the local immune response shows an increased frequency of IFN-γ-producing cells. Indeed, patients with active disease may have higher levels of IFN-γ in plasma and in sputum, suggesting that the levels of this cytokine are a reflection of the intensity of immunopathology and bacterial load in the lungs rather than evidence of a protective response (14–16). It seems likely that IFN-γ production is necessary but insufficient for protection and in the setting of concomitant M. tuberculosis disease may contribute to pathogenesis. Furthermore, the induction of IFN-γ by in vitro stimulation of PBMC may not be relevant to local protective mechanisms following aerosol exposure to viable organisms.

Simultaneous quantification of several immune functions in a single cell can now be achieved due to significant advances in multiparametric flow technology. Polyfunctional T cells show greater association with protective T cell immune responses in infectious diseases than do IFN-γ-secreting mononuclear T cells. For example, HIV nonprogressors expressed a high frequency of polyfunctional CD4+ T cells that simultaneously produced IFN-γ, tumor necrosis factor (TNF), and interleukin-2 (IL-2) (17). Similarly, antigen-specific memory T cells capable of producing these three cytokines were also protective against leishmanial infection (18). Consistent with these observations, vaccine-induced protection in mice against M. tuberculosis infection strongly correlated with a high frequency of polyfunctional CD4+ T cells (19, 20). However, in a study that monitored a cohort of BCG-immunized infants for 2 years, the correlation of polyfunctional cytokine profile with protective efficacy of BCG vaccination was not established (21). In this study, flow cytometric analysis of antigen-stimulated whole-blood samples indicated generation of multifunctional T cells in the vaccinated individuals; however, the polyfunctional profiles of these T cells did not correlate with BCG-mediated protection. In another study, T cell cytokine profile in response to the MVA85A (modified vaccinia virus Ankara expressing antigen 85A) was tested in BCG-vaccinated individuals who were given a booster dose of MVA85A. The T cell cytokine profile from these individuals indicated a significantly higher frequency of polyfunctional T cells than that for BCG vaccination alone, leading to the assumption that the booster vaccine would enhance the efficacy of BCG (22). However, despite the expansion of polyfunctional T cells with MVA85A booster immunization, the results obtained from the phase 2b trial in infants given this prime-boost strategy indicated no enhancement in protection (5). These studies suggest that polyfunctional T cells may play a necessary but not sufficient role in protection against TB.

Several investigations have suggested an IFN-γ-independent role of CD4+ T cells in mediating protective immunity to the host. Gallegos et al. showed that the adoptive transfer of antigen-specific CD4+ T effector cells could confer protection on naive hosts, independent of their ability to produce IFN-γ or TNF (23). Interestingly, using a similar approach it was shown that in vitro-differentiated BCG-specific Th17 cells mediated protection in the absence of IFN-γ (24). In another study, BCG-immunized IFN-γ−/− mice exhibited a reduction in bacterial burden on secondary challenge with M. tuberculosis infection (24, 25) but lost this ability when they were depleted of CD4+ T cells at the time of immunization (26); this study further supports the tenet that effector CD4+ T cells can mediate protection against M. tuberculosis in the absence of IFN-γ. Clearly, IFN-γ is indispensable for host protection against M. tuberculosis infection (7–10). Therefore, the requirement for IFN-γ in CD4+ T cell-mediated host resistance may be at the level of skewing CD4+ T cell differentiation toward a Th1 effector phenotype, and perhaps, IFN-γ is expendable after this stage.

Although CD4+ T cells play a predominant role in protective TB immunity, strategies that boost CD8+ function also enhance vaccine efficacy (27). In a nonhuman primate model of tuberculosis disease (28), depletion of CD8+ T cells in immunized monkeys led to reduced protection. Similarly, CD8+ T cell depletion in M. tuberculosis-infected and then antibiotic-treated monkeys led to increased susceptibility to reinfection, indicating their importance in conferring immunity in a vaccination or natural infection setting. Human CD8+CCR7+CD45RA+ effector memory T cells exhibit significant antimycobacterial activity (29), and their relevance to host protection is somewhat supported by the finding that their numbers are reduced in patients receiving immunotherapy with anti-TNF antibodies (29), a regimen that compromises M. tuberculosis immunity (30). However, whether CD8+ T cell-mediated protection is mediated by multifunctional CD8+ T cells or whether, akin to CD4+ T cells, it is also independent of IFN-γ and TNF needs to be determined.

A CASE FOR TH17 CELLS

Initial studies examining the role of IL-17 in protective immunity against M. tuberculosis found that mice lacking the ability to produce IL-17 (31, 32) and IL-17RA-deficient mice (31) were not compromised in their ability to contain M. tuberculosis growth. In contrast, another study reported that mice lacking IL-17A receptor, despite being able to control acute infection, were unable to stably maintain long-term control of M. tuberculosis infection (33). The increased susceptibility was not related to deficiencies in IFN-γ but correlated with decreased early neutrophil recruitment. However, a major caveat of this study is that the mice were infected either intratracheally or intravenously with a 100-fold-higher inoculum than the normal low-dose aerogenic route. A recent study showed that the requirement for IL-17 in host protection against M. tuberculosis was strain dependent. IL-17 was dispensable for protective immunity against the lab-adapted strain H37Rv while necessary for protection against M. tuberculosis HN878, a hypervirulent M. tuberculosis strain (34). IL-17 exerts a greater influence in vaccine-mediated protection in TB. Cooper and colleagues showed that following BCG (35) and ESAT-6 peptide (36) immunization, antigen-specific Th17 cells localized in the lungs and were critical for the recruitment of Th1 cells to the lung after M. tuberculosis challenge. Another study also found that BCG-induced Th17 cells were protective. In this study, protection was conferred even in the absence of IFN-γ (24). Indeed, other studies have reported an IFN-γ-independent mechanism of protection by Th17 cells. Mucosal vaccination with ESAT-6 peptide with LT-IB, a mucosal adjuvant, also induced a robust Th17 re-
sponse that mediated protection against *M. tuberculosis* infection in an IFN-γ-independent manner (25).

When first identified, Th17 cells were considered to be short-lived cells without the ability to generate long-term memory. Shortly thereafter, several studies showed that Th17 cells were capable of providing protection in immunization settings (37, 38) and transfer (39) models, denoting their capacity for long-term survival. It was then confirmed at the molecular level that indeed Th17 cells are long lived and can form memory cells, despite expressing markers characteristic of terminally differentiated cells (40). In fact, human Th17 cells also exhibited a long-lived effector memory phenotype, possessing a high capacity for self-renewal (41). Th17 cells preserve the molecular signature that is characteristic of T stem cell memory (TSCM) (42–44) and express the Wnt-β-catenin signaling axis (40), a pathway critical for maintaining the self-renewal potential of a cell (45, 46). Interestingly, predating Th17 and TSCM cell discovery, two studies had reported that greater protection against *M. tuberculosis* challenge infection was seen in mice that were adoptively transferred with T cells bearing a naïve phenotype (CD44<sup>+</sup>CD62L<sup>+</sup>) than in mice that were transferred with T central memory (TCM) cells (47, 48). Given our current knowledge, it is quite likely that the transferred T cells were Th17 memory cells. Together with these studies, the recent findings that Th17 cells are long lived and can mediate protection in the absence of IFN-γ suggest that the analysis of this subset of T cells is certainly warranted when evaluating new TB vaccine candidates.

OTHER IMMUNE CELLS POTENTIALLY CORRELATING WITH PROTECTION

A number of different innate cell types bearing key resemblance to T cell-like functionalities come into play during TB infection. Traditionally, these innate cells provide immediate protection before the adaptive immune response is generated and thus contribute toward early containment of the pathogen. However, a growing number of studies suggest their involvement in the recall response and protection during secondary challenge.

**NK cells.** NK cells lie at the interface of innate and adaptive immune responses and are unique in their ability to recognize and carry out a cytotoxic effector function akin to CD8<sup>+</sup> T cells, despite the lack of RAG-mediated diversity (49, 50). NK cells express only a few genes that encode a large number of different antigen-specific receptors. The role of NK cells in protection against TB has not been unequivocally proven. NK cells are present in the lungs of *M. tuberculosis*-infected mice, but their depletion does not enhance susceptibility to infection (51). However, γc<sup>−/−</sup>/RAG<sup>−/−</sup> mice that lack both NK and T cells exhibit greater susceptibility to *M. tuberculosis* infection than do RAG<sup>−/−</sup> mice, indicating a role of NK cells in host protection against *M. tuberculosis* infection (52). In vitro studies have shown that human NK cells mediate lysis of *M. tuberculosis*-infected macrophages that is dependent on expression of the NK cell-activating receptors NKP46 and NKG2D (53, 54). Mechanistically, IL-22 released by the NK cells enhanced phagolysosomal fusion and *M. tuberculosis* growth inhibition in infected macrophages (55).

Generation and long-term maintenance of NK cells in response to viral infections such as those with cytomegalovirus (CMV) and hepatitis C virus (HCV) have been reported by many investigators. These memory-like NK cells bear activating C-type lectin-like receptors such as NKG2C and depend on cytokines such as IL-12 and IL-15 for their maintenance (56, 57). Indeed, in a mouse model of CMV infection, NK cells bearing the Ly49H receptor expanded in response to infection and were maintained in the host several months postinfection. These “memory NK cells” exhibited the characteristics of memory lymphocytes by exhibiting a high rate of activation and degranulation and the ability to confer protective status on the recipient host (58). These studies emphasize the importance of investigating memory NK cells in TB vaccine assessments. In support of this, mice vaccinated with BCG demonstrated increased numbers of IFN-γ-expressing NK 1.1 cells, and their depletion led to reduced vaccination efficacy following *M. tuberculosis* challenge (59).

**γδ T cells.** γδ T cells recognize a variety of unrestricted, unprocessed, and small phosphate antigens (60, 61). In *M. tuberculosis*-infected mice, during the early phase of infection, IFN-γ- and IL-17-secreting γδ T cells with cytotoxic effector functions are recruited to the lungs (62–65). Although antibody-mediated depletion of γδ T cells in mice did not abrogate protection against BCG infection (66), expansion of Vγ2Vδ2 T cells in response to BCG vaccination and their presence in *M. tuberculosis*-specific recall response were reported in the nonhuman primate macaque model (67). In addition, human Vγ9Vδ2 T cells reduce the viability of intracellular *M. tuberculosis* via mechanisms dependent on perforin or granulysin (68–70). These data, together, indicate not only that γδ T cells are present during *M. tuberculosis* infection and BCG vaccination but that, in humans, they are capable of restricting *M. tuberculosis* growth. A study carried out in newborn pigs also showed that BCG vaccination leads to an enhanced response from γδ T cells, indicating their probable role in mediating vaccine-induced protection (71). Another study investigating characteristics of cellular response to BCG vaccination in humans found that γδ T cells expanded significantly compared to other cell types after *ex vivo* PBMC stimulation (72). Together, these studies indicate that it may be worthwhile to include functional studies of γδ T cells in the assessment of TB vaccines.

**CD4<sup>+</sup> CD8<sup>−</sup> DN T cells.** A rare subset of T cells that are CD3<sup>+</sup> TCR-αβ<sup>+</sup> but double negative (DN) for CD4 and CD8 expression expand in response to *M. tuberculosis* infection and restrict bacterial growth *in vitro* in macrophage cultures (73). Furthermore, other studies demonstrated that both a DNA vaccine cocktail (74) and BCG immunization (75) could induce protective immunity in mice lacking CD4<sup>+</sup> T cells but not in mice deficient in CD8<sup>+</sup> T cells. In contrast, the live attenuated vaccine strain mc<sub>26030</sub> was shown to induce protective immunity, equivalent to that induced by BCG, in CD4<sup>−/−</sup> mice, but in a CD8<sup>−/−</sup> T cell-independent mechanism (76). Further characterization of the T cell population mediating protection in the CD4<sup>−/−</sup> DN T cells showed that they were CD4<sup>+</sup>CD8<sup>−</sup> TCR-αβ<sup>+</sup> TCR-γδ<sup>+</sup> NK1.1<sup>−</sup> (77). Adoptive transfer of the CD4<sup>+</sup> CD8<sup>−</sup> DN T cells from vaccinated CD4<sup>−/−</sup>/CD8<sup>−</sup> mice into naive CD4<sup>−/−</sup>/CD8<sup>−</sup> resulted in significant protection against *M. tuberculosis* challenge infection (77). These enriched CD4<sup>+</sup>CD8<sup>−</sup> TCR-αβ<sup>+</sup> T cells had significantly higher mRNA levels for IFN-γ and IL-2, highlighting the need to look for correlates of protection in nontraditional CD4<sup>+</sup> and CD8<sup>+</sup> T cells. In support of this, IFN-γ-expressing DN T cells were observed in BCG-immunized children (78).

MEMORY IN THE INNATE IMMUNE COMPARTMENT

Due to its inherently nonspecific and short-lived nature, the innate immune response is not associated with the long-term mem-
ory immune response in higher vertebrates. However, studies investigating the generation of protective responses to different pathogens in invertebrate species have suggested that these hosts are able to recall prior antigenic experience, despite the lack of a specific and persistent adaptive immune response (79, 80). These protective response mechanisms involve generation of a repertoire of pattern recognition receptor molecules and the Toll pathway (79). For example, DSCAM (Down syndrome cell adhesion molecule) protects Anopheles mosquitoes against the malarial parasite Plasmodium falciparum and molecules of the Toll pathway provide protection against secondary challenge in the fruit fly Drosophila melanogaster (81–83). These evolutionarily conserved mechanisms refute the idea that innate immunity lacks specificity and is short lived (84). In higher vertebrates, pathogen-associated molecular patterns and their recognition via various pathogenic response receptors are well established and are key to providing diversity in pathogen-specific innate responses (85). However, whether this response can have an impact on long-term protection in higher vertebrates, either via generation of unique innate responses on secondary encounter or through modulation of innate responses by memory T cells, has only recently been interrogated. Another issue, of course, is whether the innate mechanisms can be expanded or enhanced through vaccination and, if so, what approaches are likely to do so.

Epigenetic reprogramming in innate cells. T and B lymphocytes undergo RAG-mediated differentiation, yielding a large repertoire of antigen-specific T and B lymphocytes, via which they then acquire effector and memory phenotypes upon antigen encounter. These memory lymphocytes persist for years and expand rapidly upon antigenic challenge, signifying their role as key targets for studying the correlates of protection in vaccination studies. However, there is emerging evidence that innate cells also undergo epigenetic reprogramming of key inflammatory genes and thereby sustain their innate activation status for long periods following the first insult (86). It is well established that plants and lower invertebrates utilize epigenetic reprogramming to maintain innate resistance in response to pathogenic challenge, but in mammals, this has not been extensively investigated (84). Indeed, BCG infection was shown to enhance the function of monocytes that lasted 3 months postinfection (87). This enhanced functional status was NOD2 dependent, and the increased gene expression of key proinflammatory cytokines was due to increased H3K4 trimethylation. Interestingly, the enhanced proinflammatory response exhibited by BCG-exposed monocytes was not limited to BCG but also included unrelated pathogens such as Candida albicans and Staphylococcus aureus (87). The authors further proposed that this epigenetic reprogramming can account for the nonspecific protection provided by BCG vaccination (84). Additional research comparing two mucosal boosting strategies has demonstrated that innate imprinting can impact the outcome of a vaccine (88). In this study, mice were first immunized parenterally with adenovirus-expressing M. tuberculosis Ag85b and then boosted by the mucosal route with either homologous vaccine or a heterologous vesicular stomatitis virus (VSV) vector expressing M. tuberculosis Ag85. Despite the induction of equivalent antigen-specific T cell responses, only mice boosted with the adenovirus vaccine showed significant protection compared to those boosted with the VSV vaccine. Interestingly, boosting with the VSV vaccine was associated with the production of IFN-β by CD11c+ b+ phagocytes, as well as downregulation of IL-12 and NOS2 (88). These observations further underscore the limitation of considering only the quantity and quality of antigen-specific T cell responses as correlates of immune protection. On the whole, these nascent studies highlight how imprinting of innate phagocytes can be utilized to enhance vaccine efficacy against TB and the need for innovative strategies to test whether vaccine candidates induce such modifications in the innate compartment.

Differential interaction of memory T cells with innate immune cells. Studies have also begun to address whether innate cells respond differently to effector and memory T cells. In this regard, the study by Strutt et al. (89) demonstrated that adoptive transfer of influenza virus–specific memory T cells to a naive recipient resulted in enhanced innate cytokine and chemokine response during secondary challenge. They observed that this enhanced innate inflammatory response was antigen specific and, importantly, independent of pathogen recognition response (PRR). Interestingly, this modulation of the innate response by memory T cells was specific to the Th1 subset but independent of IFN-γ and TNF. This study suggests that memory T cells probably interact differently with the innate component in comparison to naive T cells in a primary immune response. In a recall response, the differential interaction could lower the activation threshold of the innate cells and allow them to rapidly upregulate their antimicrobial effector response. Consistent with this possibility that effector molecules secreted by primary and secondary effector CD4+ T cells are distinct, comparative microarray analysis of the two subsets revealed approximately 450 differentially expressed genes (90). The signaling pathways between the two CD4+ T cell effector types were also found to be different (90). Another recent study also demonstrated how the innate immune response was specifically modified for enhanced protection by memory T cells in both a systemic and a mucosal model of recall response (91).

This study showed that in contrast to unvaccinated hosts, memory T cells in vaccinated hosts rapidly initiated the recruitment of innate cells, including monocytes, dendritic cells, and NK cells. Moreover, only memory T cells induced a differentiation program in the recruited innate cells, which comprised elevated expression of effector cytokines and antimicrobial pathways. However, this study found that memory T cells failed to protect if IFN-γ signaling was disrupted in the innate cells, indicating that in vaccinated hosts T cell–derived IFN-γ was key to the heightened effector response by the innate cells. Interestingly, the study that reported IFN-γ-independent modulation of innate cells investigated the effect of CD4+ T memory cells, whereas the study that found a requirement for IFN-γ to enhance effector functions of the innate cells employed a CD8+ T cell memory model. Whether it is IFN-γ or a yet-to-be-identified effector molecule from T cells that directs innate cells to mediate protection, these studies nonetheless underscore the importance of examining the effects of vaccine-induced memory T cells on innate cells. It is likely that the correlates of protection may be unearthed in the memory T cell-modulated innate cells.

**WILL STUDY OF THE NATURAL IMMUNE RESPONSE TO M. TUBERCULOSIS USING “OMICS” TECHNOLOGY PROVIDE THE ANSWER TO THE PROBLEM?**

Recent developments in high-throughput “omics” technology provide a global view of the genomic, proteomic, and metabolic status of biological systems under investigation, enabling a comprehensive view of biological processes involved in health or the
diseased state (92). Such an approach was successfully employed in predicting the immune response to yellow fever vaccine YF-17D in a cohort of healthy humans (93). In this study, the investigators used blood transcriptome profiling to monitor differences in early innate immune response to vaccination in a group of healthy volunteers and how that correlates with the magnitude of subsequent adaptive T and B cell responses. The analysis established a pattern of innate gene expression profiles in the vaccinated individuals that could predict the T cell and antibody response to 90% and 80% accuracy, respectively. A similar approach was later adopted to test the predictive value of early innate signatures for later-phase adaptive immune response to vaccine candidates against influenza (94). In this study, the investigators compared two vaccine candidates against influenza: the live attenuated influenza vaccine and trivalent inactivated influenza vaccine. In a manner similar to the study with the yellow fever vaccine, early immune response to the two influenza vaccine candidates yielded a gene signature that could be correlated with the ensuing adaptive immune response. A key marker, the calmodulin-dependent protein kinase IV (CAMKIV), identified by this strategy was successfully tested in animal models, and its role in the regulation of adaptive response to vaccination was validated (94). This wider use of “omics” technologies and system biology tools in immunology research has led to the establishment of the Human Immunology Project Consortium, which provides a platform to unite researchers engaged in large-scale data-driven research and could represent a powerful tool to use for the discovery of immune correlates of protection against TB.

Similar approaches could be used in TB to identify the correlates of protection of vaccine-mediated immunity. TB disease itself is associated with an inflammatory transcriptional blood signature absent in healthy or most latently infected individuals (95–101). There are several challenging issues in identifying the true correlates of protection against TB, including the lack of good human challenge models and lack of characterization of immune protective cells for ex vivo measurements. To obviate some of these problems, a reasonable approach would be to study the naturally induced protective immunity using “omics” technologies. The household contact (HHC) platform is a good place to start, as it offers the equivalent of a human challenge model. Household contacts of pulmonary TB cases have an intense and protracted exposure to an infectious case. The >20% that resist infection presumably are a group of heavily exposed individuals, some of whom may be protected by a vigorous innate immune response that can be characterized and potentially mimicked by vaccination. Others “self-cure” the infection, presumably by protective adaptive response. This may be indicated by reversion of tuberculosis skin test (TST) and interferon gamma release assay (IGRA) from positive to negative. This is seen in 20% of TST converters who are treated with isoniazid (INH) (102). Cohorts of household contacts also can define the susceptible phenotype, those who progress from infection to disease. The absence of candidate protective correlates in them and the presence in control populations may be of value in characterizing correlates. In fact, the INH-treated cohort may provide an immunologic profile or biomarker of "cure" of the latent focus that can be applied in a broader study of “self-cure”/protective immunity.

Based on the discussion presented here, an “omics” approach combined with single-cell technology (103) to longitudinally monitor rare cell populations, in addition to T cells, in the household contact (HHC) cohort has the potential to provide a comprehensive view of the natural immune response to M. tuberculosis in HHC. Data obtained from these immune analyses, including gene expression profiling, tetramer technology to identify antigen-specific T cells, multiparametric flow technology, and proteomics of the cohort of HHCs can then be pooled to develop predictive models and identification of biological pathways using systems biology tools (104). Modeling should also include the data related to nonspecific protection induced by BCG vaccination and demographic variation in BCG vaccination (105). The models and pathways identified can then be validated in animal models. Together, this approach has the potential to provide biomarkers predictive of a low risk of progression to disease and cure. These biomarkers would be relevant to vaccine development.

CONCLUDING REMARKS

Accumulating evidence is indicative of the deficit that exists in our current understanding of protection in primary TB or vaccination. For logical evolution of newer vaccine candidates and strategies for disease containment, it is of pivotal importance to look
ogy is a potentially promising path for uncovering the correlates of response to
system. Study of large cohorts in whom the natural immune re-
from the interaction of memory T cells with the innate immune


47. Boom WH. 1999. Gammadelta T cells and Mycobacterium tubercu-
Dieli 63.

64.

Derrick 74.


Shen

65.

295:

Sambandamurthy

Hoft

72:

tion with a DNA vaccine cocktail protects mice lacking CD4 cells against

Calmette-Guerin. J Immunol

79.

panCD: a safe and limited replicating mutant strain that protects

Jalapathy KV, Chen M, Kim J, Porcelli SA, Chan J, Morris SL.

66.

2006. Mycobacterium tuberculosis DeltaRD1 Delta-

panCD: a safe and limited replicating mutant strain that protects immunocompetent and immunocompromised mice against experi-


571–476.

67.


68.

2000. Vgamma9/Vdelta2 T lymphocytes re-

duce the viability of intracellular Mycobacterium tuberculosis. Eur J Immunol

505.4662–4669. http://dx.doi.org/10.4049/jimmunol.177.7.4662.

69.

Dieli

70.

2004. Granzyme A produced by gamma(9)delta(2) T cells induces human


252.0099.

71.


81:


311:


jimmunol.0903332.

82.


83.


84.


panCD: a safe and limited replicating mutant strain that protects immunocompetent and immunocompromised mice against experi-


85.


86.


